AMENDMENT

Please amend the application without prejudice, without admission, without surrender of subject matter, and without any intention of creating any estoppel as to equivalents, as follows.

In the Specification

Please amend the paragraph beginning on page 33, line 15 as follows:

Further homologues of known mammalian Notch ligands may be identified using standard techniques. By a "homologue" it is meant a gene product that exhibits sequence homology, either amino acid or nucleic acid sequence homology, to any one of the known Notch ligands, for example as mentioned above. Typically, a homologue of a known Notch ligand will be at least 20%, preferably at least 30%, identical at the amino acid level to the corresponding known Notch ligand over a sequence of at least 10, preferably at least 20, preferably at least 50, suitably at least 100 amino acids, or over the entire length of the Notch ligand. Techniques and software for calculating sequence homology between two or more amino acid or nucleic acid sequences are well known in the art (see for example ~~http://www.ncbi.nlm.nih.gov~~ the website for the National Center for Biotechnology Information and Ausubel *et al.*, Current Protocols in Molecular Biology (1995), John Wiley & Sons, Inc.)

Please amend the figure descriptions on page 15, lines 4-30, as follows:

~~Figures~~ Figure 4 shows a schematic representation of ~~the assays of~~ CD4+ T-cells cultured in plates coated with anti-CD3 antibody and Delta-Fc, and resuspended with anti-CD28 antibody, as described in Examples 1 to [[9]] 3;

Figure 5 shows a schematic representation of ~~the assays of~~ a CHO-N2 cell cultured in plates coated with Delta-Fc, and transfected with a Luciferase reporter plasmid, as described in Examples [[1]] 2 to [[9]] 8;

Figure 6 shows a schematic representation of ~~the assays of~~ a co-culture assay comprising CHO-N2 cells transfected with a Luciferase reporter plasmid and plated, and CHO-Delta or A20-Delta cells, as described in ~~Examples 1 to 9~~ Example 8;

Figure 7 shows ~~the results of~~ ELISA measurements of cytokine production with and without the addition of soluble Fc-delta, as described in Example 3;

Figure 8 shows ~~the results of~~ HES-1 expression relative to 18S rRNA expression with and without Fc-delta, as described in Example 4;

Figure 9 shows ~~the results of~~ ELISA measurements of cytokine production under polarizing conditions, as described in Example 5;

Figure 10 shows ~~the results of~~ cytokine production and compares soluble Fc-delta with plate-bound Fc-delta against control, as described in Example 6;

Figure 11 shows ~~the results of~~ cytokine production and cell proliferation in cells experiencing secondary stimulation, as described in Example 7;

~~Figure 12 shows~~ Figures 12A-12D show the results of sample assays using the CHO-Notch2-10xCBF1-Luc reporter cell line with (A) immobilised human Delta-1/Fc fusion protein, (B) plate-immobilised mouse Delta-1/Fc fusion protein, (C) CHO/CHO-human Delta1 co-cultured cells and (D) A20/A20-mouse Delta1 co-cultured cells as actives against corresponding controls, as described in Example 8;

Figure 13 shows ~~the results of~~ measurements of cytokine production induced by Delta-Fc coated beads in T-cells, as described in Example 10;

Figures 14A and 14B show ~~the results of~~ IL-10 production in the presence of mouse or human Delta1 beads in T-cells from (A) Donor 1 and (B) Donor 2, as described in Example 11;

Figures 15A and 15B show ~~the results of~~ IL-5 production in the presence of mouse or human Delta1 beads in T-cells from (A) Donor 1 and (B) Donor 2, as described in Example 11;

Figure 16 shows ~~the results of~~ IL-10 production in the presence of mouse Delta1, as described in Example 11;

Figure 17 shows ~~the results of~~ IL-5 production in the presence of mouse Delta1 beads, as described in Example 11;

Figure 18 shows ~~the results of~~ cytokine production in the presence of mouse Delta1-Fc beads in T-cells, as described in Example 11;

Figure 19 shows ~~the results of~~ IL-10 production in the presence of Delta1 beads in T-cells at varying bead-to-cell ratios, as described in Example 12;

Figures 20A and 20B show ~~the results of~~ cytokine production in the presence of mouse Delta1-Fc beads in (A) Unseparated CD4+ T-cells and (B) Memory CD4+CD45RO+ T-cells, as described in Example 13;

Figure 21 shows ~~the results of~~ IL-10 production in the presence of Delta-Fc in un-polarised cells, Th1 polarised cells, and Th2 polarised cells, as described in Example 14;

Figures 22A and 22B illustrate ~~the results of~~ (A) a schematic representation of the protocol for activating with Delta alone and (B) a Venn diagram showing numbers of genes with increased expression in response to Delta activation alone, as described in Example 15;

Figure 23 shows ~~the results of~~ a time-course expression profile of Delta-mediated activation of gene expression in Jurkat T-cells, as described in Example 15;

Figures 24A and 24B illustrate ~~the results of~~ (A) a schematic representation of the protocol for activating with both Delta and anti-CD3/CD28 activation, and (B) a Venn diagram showing numbers of genes with increased expression in response to Delta activation in combination with anti-CD3/CD28 activation, as described in Example 15;

Figures 25A and 25B show ~~the results of~~ a time-course expression profile for (A) Delta activation in combination with anti-CD3/CD28 activation and (B) Delta activation alone, as described in Example 15;

Figures ~~26A, 26B and 26C~~ 26A-26C show ~~the results of~~ gene expression in response to Delta activation with and without T-Cell Receptor Stimulation for (A) Cip1-interacting zinc finger protein, (B) UMP-CMP kinase, and (C) Helicase, as described in Example 15;

~~Figures~~ Figure 27 shows the results of transient reporter assays conducted with Jurkat/Notch2 clones with or without immobilised hDLL1-Fc and with or without PMA/ionomycin, as described in Example 16;

~~Figures~~ Figure 28 shows ~~the results of~~ the reporter assay dose response to the concentration of plate-bound hDLL1-Fc in Jurkat/Notch2 clones, as described in Example 16;

Figure 29 shows ~~the results of~~ a reporter assay dose response to ionomycin concentration, as described in Example 17;

~~Figures 30A and 30B show the results of~~ Figure 30 shows the effects of treating Jurkat cells transfected with Human Notch1 intracellular domain and reporter constructs with anti-CD3/CD28 or PMA/ionomycin assessed by fold activation compared to absence of Notch IC (Expt 1) and luminescence (Expt 2), as described in Example 18;